

BIOSYNTHETIC STUDIES OF MARINE LIPIDS 15.¹

CONVERSION OF PARKEOL (LANOSTA-9(11),24-DIEN-3 β -OL) TO

14 α -METHYLCHOLEST-9(11)-EN-3 β -OL IN THE SEA CUCUMBER *HOLOTHURIA ARENICOLA*

M. Lucinda Cordeiro, Russell G. Kerr and Carl Djerassi*

Department of Chemistry, Stanford University, Stanford, CA 94305

Abstract: Tritium labeled parkeol (2), but not lanosterol (4) or cycloartenol (3), was transformed by the sea cucumber *Holothuria arenicola* into 14 α -methyl-5 α -cholest-9(11)-en-3 β -ol (5), thus implying that squalene-2,3-oxide cyclizes directly to parkeol (2).

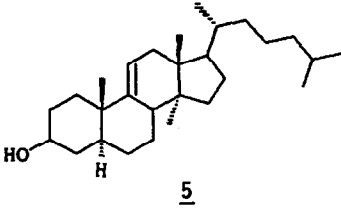
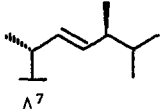
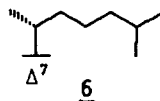
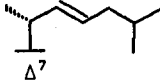
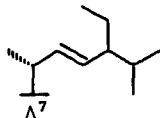
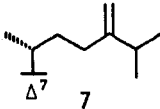
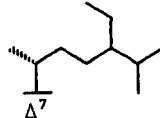
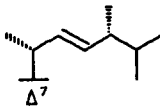
Biosynthetic work² in this laboratory has focused on the genesis of the unusual side chains found in sponge sterols, but recently attention has also been paid to the earlier stages of sponge sterol biosynthesis.³ In this connection we have re-examined some earlier work on the sea cucumber *Stichopus californicus*⁴ found near Stanford University's Hopkins Marine Station in Monterey Bay. In addition to conventional Δ^7 -sterols (cf. Table 1), we isolated from the free sterol fraction (without prior saponification or acid hydrolysis) the biosynthetically intriguing 14 α -methylcholest-9(11)-en-3 β -ol (5).⁵ This same sterol, along with conventional Δ^7 -sterols (see table 1), was also isolated from several other sea cucumbers including the Caribbean *Holothuria arenicola*, with which this biosynthetic study is concerned.

There are few other instances of the occurrence of the rare $\Delta^{9(11)}$ -14 α -methyl nuclear substitution pattern in nature: 14 α -methyl-5 α -ergosta-9(11),24(28)-dien-3 β -ol from a member of the Cucurbitaceae;^{7a} 4 α ,14 α -dimethyl-5 α -cholest-9(11)-en-3 β -ol from a member of the Solanaceae;^{7b} and 14 α -methyl-5 α -cholest-9(11)-en-3 β -ol (5) from the sea cucumbers *Psolus phantapus*^{7c} and *Psolus fabricii*.^{7d} Goad *et al.*^{7d} speculated that this latter sterol (5) may arise by cyclization of squalene-2,3-oxide to an intermediate cation 1, which subsequently gives rise to both lanosterol (3) and parkeol (2). They further suggest that 3 is the immediate precursor of the major sterol lathosterol (6), which possesses the conventional sterol skeleton (without any methyl groups at C-4 and C-14), whereas parkeol (2) is metabolized via loss of the two C-4 methyl groups to the 14-methylated sterol (5). In the latter case, loss of the C-14 methyl group presumably does not occur due to lack of activation by an 8,9 double bond.⁸ By contrast, Akhila *et al.*⁹ suggested that 5 α -stigmast-9(11)-en-3 β -ol, a $\Delta^{9(11)}$ sterol lacking the 14 α -methyl group arises by cyclization of squalene oxide to parkeol (2), possibly via prior formation of cycloartenol (4).

In order to distinguish between these various metabolic possibilities, we fed [24-³H]-labeled¹⁰ lanosterol (3), cycloartenol (4) and parkeol (2) to *Holothuria arenicola*,¹⁰ and examined the incorporation of radioactivity among its sterols (Table 1). In each instance,

approximately 20 μ Ci (44×10^6 dpm) of precursor in 1 mL of 70% ethanol was administered to the Holothurian by injection into the coelomic cavity and the animal maintained for 7 days in an aquarium in circulating seawater with aeration before sacrifice. The sterols were purified by normal phase HPLC (6% ethyl acetate in hexanes) followed by reverse phase HPLC using methanol and then 30 mM AgNO_3 in 97% methanol. The results were quite unambiguous. No radioactivity was found in any of the eight sterols listed in Table 1 when labeled lanosterol (3) or cycloartenol (4) were fed, although 7.7×10^5 dpm and 6.2×10^5 dpm were noted in the lanosterol and cycloartenol fractions, respectively, showing that the precursors were taken up by the sea cucumber. When parkeol (2) was administered, significant radioactivity (45,500 dpm after repeated purification to constant radioactivity using cold 5 as carrier during HPLC) was encountered in 14 α -methyl-5 α -cholest-9(11)-en-3 β -ol (5), while all the other seven sterols (Table 1) were cold. We conclude that (a) the $\Delta^{9(11)}$ -14 α -methyl sterol (5) is the only *bona fide* sterol metabolite in the sea cucumber and (b) that the remaining Δ^7 -14 α -desmethyl sterols (e.g. 6 or 7) are dietary constituents or late metabolites (e.g. side chain alkylation) of a dietary precursor which already possesses a standard 4,4,14-desmethyl sterol skeleton.

TABLE 1. FREE STEROLS IN *HOLOTHURIA ARENICOLA*

Sterol ^a	Relative Abundance (%)	Sterol ^a	Relative Abundance (%)
	7.8		6.4
<u>5</u>			22.5
	3.8		14.9
	24.1		12.4
	6.8	unidentified trace sterols	1.3

^a Δ^7 denotes the 5 α -androst-7-en-3 β -ol nucleus.



Acknowledgements: Financial support by the National Institutes of Health (grants no. GM06840 and GM28352) is gratefully acknowledged.

1. For preceding paper, see J.-H. Cho, I.L. Stoilov, J.E. Thompson, C. Silva and C. Djerassi, J. Org. Chem., submitted for publication.
2. (a) C.A.N. Catalan, J.E. Thompson, W.C.M.C. Kokke and C. Djerassi, Tetrahedron 41 (1985) 1073; (b) J.R. Proudfoot, C.A.N. Catalan, C. Djerassi, D. Sica and G. Sodano, Tetrahedron Lett. 27 (1986) 423; (c) I.L. Stoilov, J.E. Thompson and C. Djerassi, Tetrahedron 42 (1986) 4147; (d) I.L. Stoilov, T.G. Back, J.E. Thompson and C. Djerassi, Tetrahedron 42 (1986) 4156; (e) I.L. Stoilov, J.E. Thompson and C. Djerassi, Tetrahedron Lett. 27 (1986) 4821; (f) I.L. Stoilov, J.E. Thompson, J.-H. Cho and C. Djerassi, J. Org. Chem. 108 (1986) 8235.

3. R.G. Kerr, I.L. Stoilov, J.E. Thompson and C. Djerassi, manuscript in preparation.
4. Y.M. Sheikh and C. Djerassi, Tetrahedron Lett. (1977) 3111.
5. This sterol was identified by comparison of ^1H NMR and GC-MS data with literature values: L.J. Goad, F.-X. Garneau, J.W. ApSimon and M. Girard, Tetrahedron Lett. (1985) 3513. The earlier reported (ref. 4) isolation of holotoxinogenin could not be replicated.
6. Further work on *Stichopus californicus* and other holothurians from Australia and Puerto Rico is underway in our laboratory.
7. (a) T. Akihisa, T. Tamura and T. Matsumoto, Phytochemistry 26 (1987) 2412; (b) T. Itoh, T. Ishi, T. Tamura and T. Matsumoto, Phytochemistry 17 (1978) 971; (c) K.B.M. Ambia, L.J. Goad, S. Hkycko, F.-X. Garneau, J. Belanger and J.W. ApSimon, Comp. Biochem. Physiol. 86B (1987) 191; (d) L.J. Goad, F.-X. Garneau, J.L. Simard, J.W. ApSimon and M. Girard, Comp. Biochem. Physiol. 84B (1986) 189.
8. For leading references, see W.R. Nes and M.L. McKean, in "Biochemistry of Steroids and other Isopentenoids," University Park Press: Baltimore, 1977, chapter 9.
9. A. Akhila, M.M. Gupta and R.S. Thakur, Tetrahedron Lett. (1987) 4085.
10. 24-Oxolanosterol acetate was synthesized by epoxidation of lanosterol acetate and subsequent opening of the epoxide with boron trifluoride etherate (Y. Sato and Y. Sonoda, Chem. Pharm. Bull. 29 (1981) 356), and then converted to ^3H labeled lanosterol by NaB^3H_4 reduction and POCl_3 dehydration (K.H. Raab and W.R. Nes, Biochem. Biophys. Acta, 152 (1968) 742), followed by treatment with LiAlH_4 . Labeled cycloartenol was synthesized in a similar manner using 24-oxocycloartenol acetate. Parkeol was prepared by isomerization of 24-oxolanosterol with 10% trifluoroacetic acid in benzene at room temperature for 72 hrs. The ^3H label was then introduced at C-24 as described above.
11. G.F. Gibbons, L.J. Goad, T.W. Goodwin and W.R. Nes, J. Biol. Chem. 246 (1971) 3967.
12. A.G. Smith and L.J. Goad, J. Biochem. 146 (1975) 35.

(Received in USA 13 January 1988)